

Work Package 3: Activity-based bioprospecting for enzymes



Meeting #3

General Assembly – online – November 14 2022



Project funded by the European Union's Horizon 2020
Research and Innovation Programme under grant agreement No [101000327]



Work package number ⁹	WP3	Lead beneficiary ¹⁰	3 - BANGOR
Work package title	Activity-based bio-prospecting for enzymes		
Start month	1	End month	36

Objectives

Entirely novel enzymes, which have no homologues in databases or with homology to known enzymes lower than ~20%, will escape the computational pre-screening in WP2. In order to circumvent this problem, WP3 combines three main pillars: bio-resources, technical capabilities for bio-resource handling and management, and activity-based multi-screens and Next Generation Sequencing (NGS), with a major objective: To screen for novel enzymes which, because their novelty, could not have been predicted using the BLAST, HMM and computational screens of WP2. To this end, the consortium has established a large collection of environmentally, geographically and taxonomically diverse bio-resources: 1. Cultured microbial isolates from (non)-extreme environments, many representing novel lineages; 2. Expression libraries from DNA from uncultivable microbial communities. 3. Genome sequences of cultivable microbial isolates; 4. Shotgun sequences of microbial communities; 5. Enzymes available in expression systems. The consortium has also at its disposal technical facilities and tools for: 1. Microbial handling and cultivation, including extremophiles, bioprospecting and sampling; 2. Handling and cloning DNA from uncultivable microorganisms; 3. High throughput screening supported by robotic and single cell manipulation workstations; 4. Multiple complementary analytics; 5. DNA sequencing (Illumina MiSeq and Oxford Nanopore instruments); 6. Bioinformatics analysis.

Description of work and role of partners

WP3 - Activity-based bio-prospecting for enzymes [Months: 1-36]

BANGOR, CSIC, UHAM, UDUS, IST ID, CNR

In WP3 we propose 3 major tasks, through which we will implement sophisticated activity-based platforms to exploit available and new ad hoc bio-resources for entirely novel enzymes demanded by the detergent, textile and cosmetic sectors.

Task 3.1 Exploitation of the FuturEnzyme bio-resource collections M1-M24

Task Lead Partner – IST-ID

Participants: CSIC, UHAM, UDUS, BANGOR, CNR

Task 3.2 Sampling extreme environments for generating new microbial bio-resources M6-M30

Task Lead Partner – CNR

Participants: IST-ID

Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36

Task Lead Partner – BANGOR

Participants: CSIC, UHAM, CNR

Participation per Partner

Partner number and short name	WP3 effort
1 - CSIC	5.00
3 - BANGOR	17.00
4 - UHAM	10.00
5 - UDUS	4.00
6 - IST ID	21.00
7 - CNR	18.00
Total	75.00

List of deliverables

Deliverable Number ¹⁴	Deliverable Title	Lead beneficiary	Type ¹⁵	Dissemination level ¹⁶	Due Date (in months) ¹⁷
D3.1	Bio-resources prepared and exchanged	3 - BANGOR	Other	Confidential, only for members of the consortium (including the Commission Services)	2
D3.2	Standard assays, analytics and calculations for monitoring enzymatic performance	4 - UHAM	Report	Confidential, only for members of the consortium (including the Commission Services)	2
D3.3	Set of 100 best clones, 10 isolates, and 10 enzymes shortlisted for sequencing or transfer to WP2	6 - IST ID	Other	Confidential, only for members of the consortium (including the Commission Services)	10
D3.4	Sequence, activity, and stability datasets from best positive bio-resources	3 - BANGOR	data sets, microdata, etc	Confidential, only for members of the consortium (including the Commission Services)	18
D3.5	Set of new bio-resources to screen or sequence	7 - CNR	Other	Confidential, only for members of the consortium (including the Commission Services)	24
D3.6	Complete set of positive naïve screened enzymes and sequences and their datasets	3 - BANGOR	Report	Confidential, only for members of the consortium (including the Commission Services)	32

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- A list of bio-resources available within the consortium have been prepared and exchange (D3.1 – month M2)
 - All partners implicated

D3.1. BIO-RESOURCES PREPARED AND EXCHANGED



ENZYMES AVAILABLE

1353 entries, representing highly diverse enzymes relevant to FuturEnzyme, available in expression systems, from single (meta)genomes; the enzymes have been isolated and characterized for purposes others than those in FuturEnzyme, and will be now screened with project-relevant substrates and conditions.



ISOLATES AVAILABLE

1387 entries, representing psychrophilic, mesophilic, thermophilic, hyper-thermophilic, thermo-acidophilic, alcaliphilic, extreme halophilic, obligate anaerobic and facultative (micro)aerobic sulphur-respiring microorganisms. The collection includes strains growing at temperatures from 0° to 92°C, pH from 1.5 to 10.0, salinity up to 490 g/L, and pressure up to 50 Mpa.



ISOLATES WITH GENOMES AVAILABLE

197 entries, representing genomes from isolates representing lineages of (non)-extremophiles growing from 0 to 92°C, pH from 1.5 to 9.0, salinity up to 492 g/L, pressure up to 50 MPa.



METAGENOMIC LIBRARIES

28 entries, representing DNA material from communities inhabiting extreme environments (low pH from 1.1 to 4.4; high pH of 9.3-9.6; high salinity from 200 to 490 g/L; pressure up to 300 MPa; temperature up to 98°C) and non-extreme environments, including contaminated sites (close to neutral pH, low to moderate salinity (up to 50 g/l), temperatures from 4 to 30°C, up to 10.1 MPa).



ENRICHMENT CULTURES

41 entries, derived from samples originated from multiple locations and representing enriched microorganisms of at least 16 different genera.



ISOLATES WITH PROVEN ACTIVITY

55 entries.



SHOTGUN METAGENOME SEQUENCES

61 entries, corresponding to at least 16 different types of extreme and non-extreme environments.

These QR codes are confidential and available within the FuturEnzyme consortium. In order to increase the security, they have been blocked with a password (FuturEnzyme€01/06/2021). They will also be included in the private area of the FuturEnzyme website (www.futurenzyme.eu), in the section Shared material. This private area that serves as a repository for the project is accessible to the members of the consortium through user and password.

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- A number of assays for functional screens have been defined and shared (D3.2 – Month 2)
 - All partners implicated



D3.2. STANDARD ASSAYS, ANALYTICS AND CALCULATIONS FOR MONITORING ENZYMATIC PERFORMANCE



18x Protocols for DETERGENT APPLICATIONS

- 4x pH shift liquid protocols for quantifying esterase-lipase activity
- 3x Liquid protocols for quantifying esterase-lipase activity with chromogenic esters
- 2x Liquid protocols for quantifying esterase-lipase activity with non-chromogenic esters
- 9x Agar plate protocols



23x Protocols for TEXTILE APPLICATIONS

- 7x Agar plate polyesterase screening assays
- 2x Agar plate protease protocols
- 1x Agar plate cellulase protocol
- 1x Agar plate oxidoreductase protocol
- 12x Liquid protocols



7x Protocols for COSMETIC APPLICATIONS

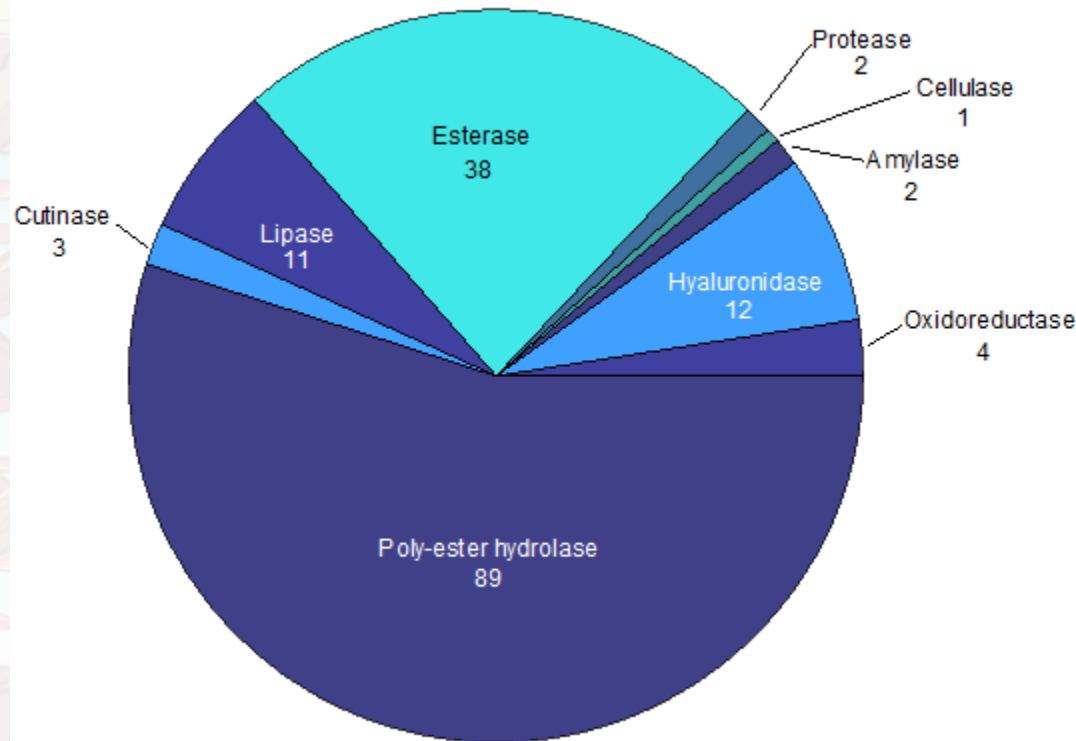
- 4x Liquid colorimetric assays for hyaluronidase activity
- 1x Liquid analysis of hyaluronic acid oligosaccharides by HPAEC-PAD
- 1x Liquid analysis of hyaluronic acid oligosaccharides by SEC-ELSD
- 1x Agar assay for the determination of hyaluronidase activity

**48 common and
standardised
protocols**

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- A list of best clones, isolates and enzymes have been prepared and shortlisted for sequences (D3.3 – month M10)
 - All partners implicated

D3.3. 100 Best clones, 10 isolates, and 10 enzymes shortlisted for sequencing or transfer to WP2



The 154 prospects (up to 162, noticing that some isolates present different activities, so most probably several enzymes) have been settled apart out of 120 genomes from isolates, metagenomes from 47 microbial communities, 1200 microbial strains, 30 metagenome libraries and 500 enzymes (as mentioned in Task 3.1, Grant Agreement, Annex 1, part A).

Distribution by activity of the enzymes and microorganisms selected in this deliverable

Isolates/Microorganisms

			Number of isolates with activity/ies	First priority for (industrial partner/s)	Second priority for (industrial partner/s)
Oxidoreductase			3	Schoeller	Henkel
Hydrolase	Glucosidase	Hyaluronidase	10	Evonik	
		Amylase	1	Scholler	Henkel
	Peptidase	Protease	1	Scholler	Henkel
	Ester-hydrolase	Esterase	4	Henkel/ Schoeller	
		Lipase	8	Henkel/ Schoeller	
		Cutinase	3	Henkel/ Schoeller	

Enzymes

				Number of enzymes with activity	First priority for (industrial partner/s)	Second priority for (industrial partner/s)
EC 1	Oxidoreductase	Laccase, Cu-oxidase		1	Schoeller	Henkel
EC 3	Hydrolase	Glucosidase	Hyaluronidase	2	Evonik	
			Amylase	1	Scholler	Henkel
			Cellulase	1	Schoeller	
		Peptidase	Protease	1	Scholler	Henkel
		Ester-hydrolase	Esterase	34	Henkel/ Schoeller	
			Lipase	3	Henkel/ Schoeller	
			Poly-ester hydrolase	89	Schoeller	Henkel

D3.3. 100 Best clones, 10 isolates, and 10 enzymes shortlisted for sequencing or transfer to WP2



The following QR code directs to the full list of **candidate isolates**



The following QR code directs to the full list of **candidate enzymes**

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- Data set of sequence, activity and stability from best positive bio-resources (D3.4 – month M18) ***to be finalised now!***

UDUS

Previously
reported

• WP3

➤ Task 3.1 Exploitation of the FuturEnzyme bio-resource collections

✓ A set of 85 esterases and lipases including 16 cutinase-like enzymes from previous projects collected



	1	2	3	4	5	6	7	8	9	10	11	12	
A	LCC WCCG	Psab PE-H	Abo_LipA (CE02)	Paes TB045 (CE15)	lipA	Est24c11	TBEc350	Hyd8c31		Aku GD5L1 (CE17)		Hyd20c2	A
B	LCC	Paby PE-H	Hyd18c13	Abo_LipC (CE09)	EV	TBEcIH8	TBEc321	Hyd18c8	Hyd4c6	Est8c8		Lip10c11	B
C	PETase	Ppac PE-H	TBEc157	Hyd13c2	Aku Est3 (CE20)		Est9c19	Lip3c12	Hyd3c14	Abo_LipE (CE10)		Est20c28	C
D	Plit PE-H	Poce PE-H	TBEc310	Abo_LipD (CE07)	Est9c28	Hyd10c19	TBEc314	Aku Est2 (CE19)	Abo_LipG (CE11)	Abo_LipI (CE12)		Est13c9	D
E	Pbau PE-H	Paes PE-H Y250S	TBEc304	Hyd33c4	Lip1c6	Est29c9	Hyd20c11	Paes TB035 (CE13)	1,4-D#003	Hyd22c10		Est30c5	E
F	Ppel PE-H	Paes PE-H (CE16)	CycTB025	Abo_Est3 (CE03)	MHETase	Est51c6	Aku Est1 (CE18)	TBEc305	Hyd23c15	TBEc308	EV	EstP	F
G	POIL-1 PE-H	Pxin PE-H	Hyd7c19	EV	Dim-008 (CE01)	Paes TB037 (CE14)	Est16c36	ED30	Est24c4	Hyd19c35	Est65c2		G
H	Abo_Est7 (CE05)	Paes TB072 (CE24)	Dim004	Abo_Est1	Paes TB040 (CE22)	FScut	PaesTB074 (CE25)	Dim001	Paes TB081 (CE23)	Abo Est2 (CE04)	Paes TB001 (CE21)	Abo Est12 (CE08)	H

Table of ready to use UDUS/UHAM esterases & lipases at UDUS for FuturEnzyme activities

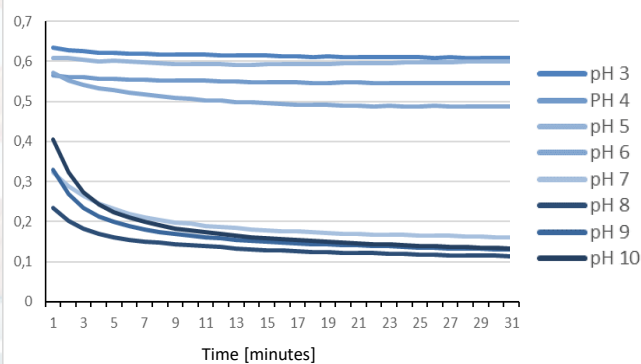
UDUS

Previously
reported

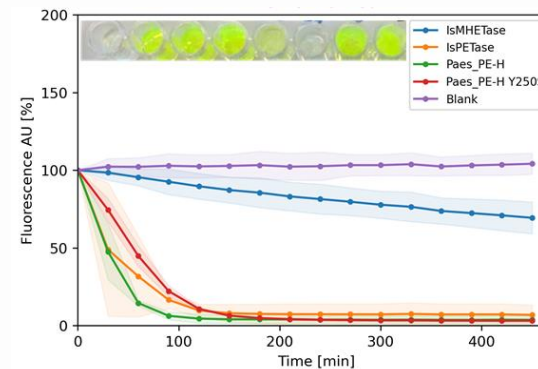
• WP3

- Task 3.1 Exploitation of the FuturEnzyme bio-resource collections
- ✓ A set of 88 esterases and lipases including 16 cutinase-like enzymes from previous projects collected
- ✓ MTP assay development

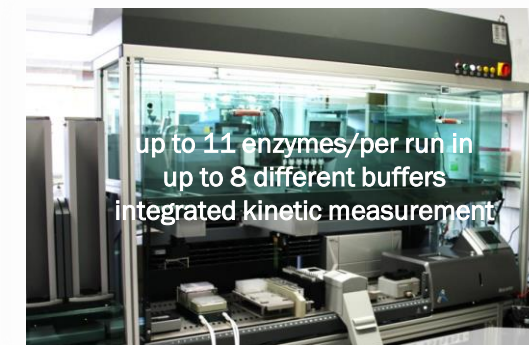
Turbidity measurements for
cutinase characterization



pH indicator assay for hydrolysis of
solid substrates



Implemented automated
characterization



UDUS

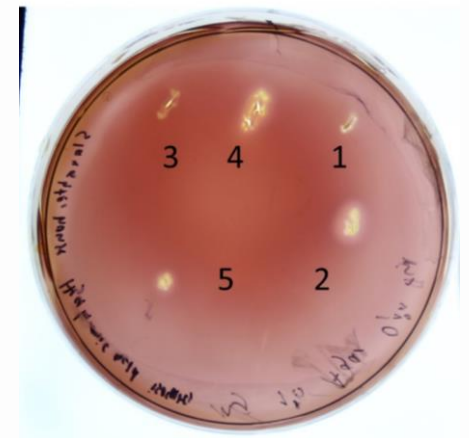
Previously
reported

- WP3

- Task 3.1 Exploitation of the FuturEnzyme bio-resource collections

- ✓ Enrichment cultures of slaughterhouse metagenome bank with hyaluronic acid
 - Several strains with *in silico* put. hyaluronic acid lyases
 - Mainly from clade of γ-gamma proteobacteria,
e.g., *Proteus* sp. or *Raoultella* sp.

M9 medium
+0.1% hyaluronic acid



UDUS

• WP3 Deliverable 3.3 Resources



	Source	Id-status	Genome status	Enzyme candidates
isolates showing hyaluronic acid hydrolysis				
Proteus sp.	Slaughterhouse drain	Partial 16S rDNA sequence	Not sequenced, Sequence of related strains available	<i>In silico</i> : Chondroitin lyase Put. Hyaluronic acid AC Lyase
Isolate Hyal_hyal_UDUS 2 (Raoultella sp.)	Slaughterhouse drain	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Next step: <i>in silico</i> candidate enzyme identification (BSC?)
Isolate Hyal_hyal_UDUS 3 (Raoultella sp.)	Slaughterhouse drain	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Next step: <i>in silico</i> candidate enzyme identification (BSC?)
(Isolate Hyal_hyal_UDUS 4) (Spirosoma)	Slaughterhouse drain	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Next step: <i>in silico</i> candidate enzyme identification (BSC?)
strains exceptionally enriched in esterases				
Halopseudomonas aestusnigri	Oil polluted coast (spain)	Type strain	draft	2 Polyesterases (1 confirmed), 12 confirmed additionally esterases
Halopseudomonas litoralis	Coastal waters (spain)	Type strain	Closed genome available.	2 Polyesterases (1 confirmed)
Halopseudomonas oceani	Deep Sea	Type strain	draft	2 Polyesterases (1 confirmed)
Halopseudomoans bauzanensis	Polluted industrial site soil (italy)	Type strain	draft	2 Polyesterases (1 confirmed)

UDUS

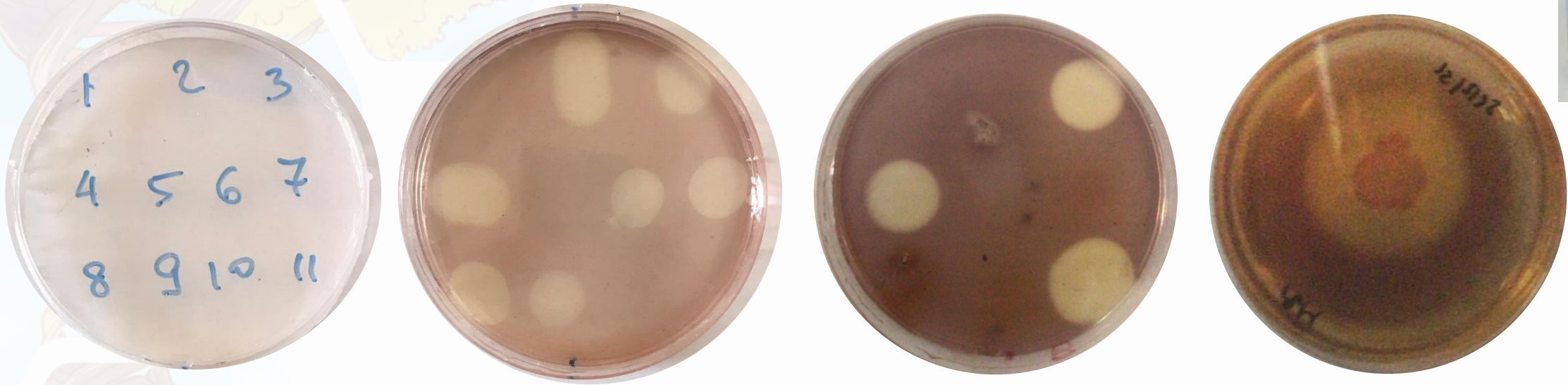
• WP3 Deliverable 3.3 Resources



	Source	Id-status	Genome status	Enzyme candidates
isolates showing polyester hydrolysis				
PEH_PUE UDUS1	Deep Sea sediment	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Candidates identified by HMM (UHAM). Next step:cloning
PEH_PUE UDUS2	Deep Sea sediment	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Candidates identified by HMM (UHAM). Next step:cloning
PEH_PUE UDUS3	Deep Sea sediment	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Candidates identified by HMM (UHAM). Next step: cloning
PEH_PUE UDUS4	Deep Sea sediment	Partial 16S rDNA sequence	Sequencing finished, draft (by Bangor)	Candidates identified by HMM (UHAM). Next step: cloning
PEH_PUE UDUS5	Deep Sea sediment	Type strain	Sequencing finished, draft (by Bangor)	Candidates identified by HMM (UHAM). Next step: cloning

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- Two hyaluronic-acid degrading isolates have been identified by CNR, when using EVO hyaluronic acid as substrate for screening



- Halorhabdus* sp. SivX81 (genome sequenced)
- Vibrio alginolyticus* from anoxic sediments of meromictic brackish Lake Faro, Messina

List of hydrolytic halo- and halonatronoarchaea

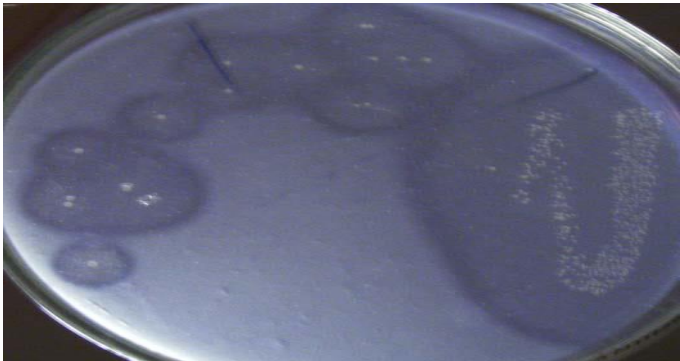
Strain	Lakes	Affiliation	Substrate	Activity	Growth
H-hyl	Cl	Halobacteria (new genus?)	Hyaluronic acid (HA)	+	+
RMX81	Cl	Halorhabdus sp.	Coco oil, xylan. HA	+	+
RMX62	Cl	Halorhabdus uthaensis	Coco oil, skim milk, xylan, HA	+	+
Siv8X	Cl	Halorhabdus uthaensis	Coco oil, skim milk, xylan HA	+	+
HArcel-Eu2	Cl	Halomicrobium sp.	Hyaluronic acid, cellulose	+	+
HArcel2**	Cl	Halosimplex sp.	Cellulose	+	+
HArcel3**	Cl	Halomicrobium sp.	Cellulose	+	+
Harc-L1	Cl	Halobacteria (unidentified)	Olive oil	+	+
Harc-L2	Cl	Halobacteria (unidentified)	Olive oil	+	+
BNX81	Cl	Halococcoides cellulosivorans	Cellulose, xylan HA	+	+
LCL711		Halorhabdus sp.	Xylan, hyaluronic acidHA	+	+
AB-hyl1	SL	Paracoccus sp.	Hyaluronic acid (HA)	+	+
AArcel7	SL	Natrarchaeobius sp.	Hyaluronic acid (HA)	+	-
AArc-St1-1*	SL	Natranaeroarchaeum aerophilum	Hyaluronic acid (HA)	+	-
AArc-L1	SL	Natrarchaeobaculum aegyptiacus	Olive oil	+	+
AArc-L2	SL	Natronolimnohabitans innermongolicus	Olive oil	+	+
AArc-LBj	SL	Halobacteria (unidentified)	Olive oil	+	+

S - soda lakes; Cl - chloride lakes;

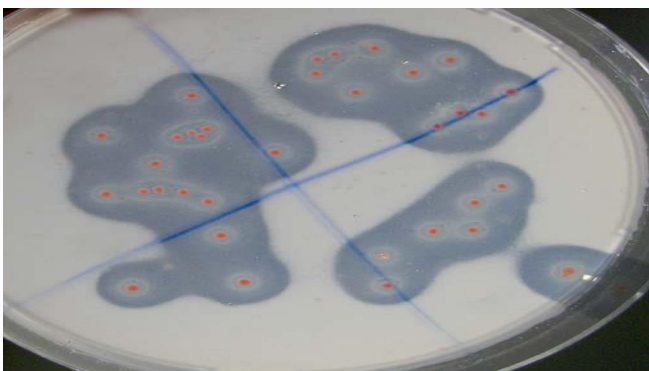
* - extremely high carotenoid content - suitable for production?

** - for genomes

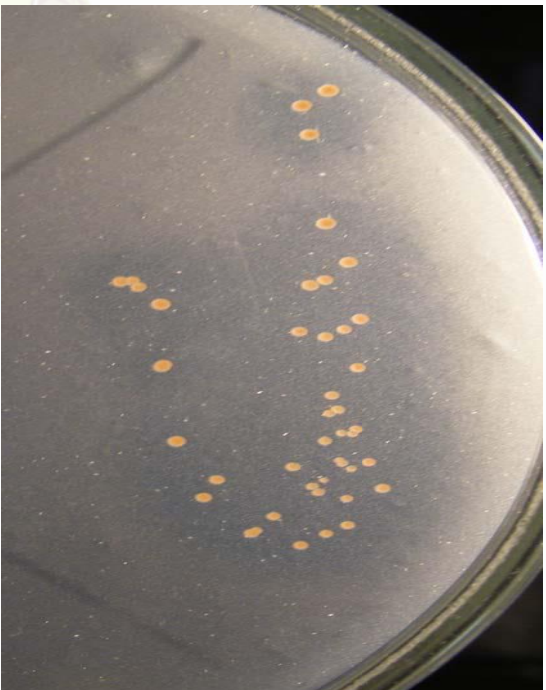
Bold – genome sequenced



AArc-L1



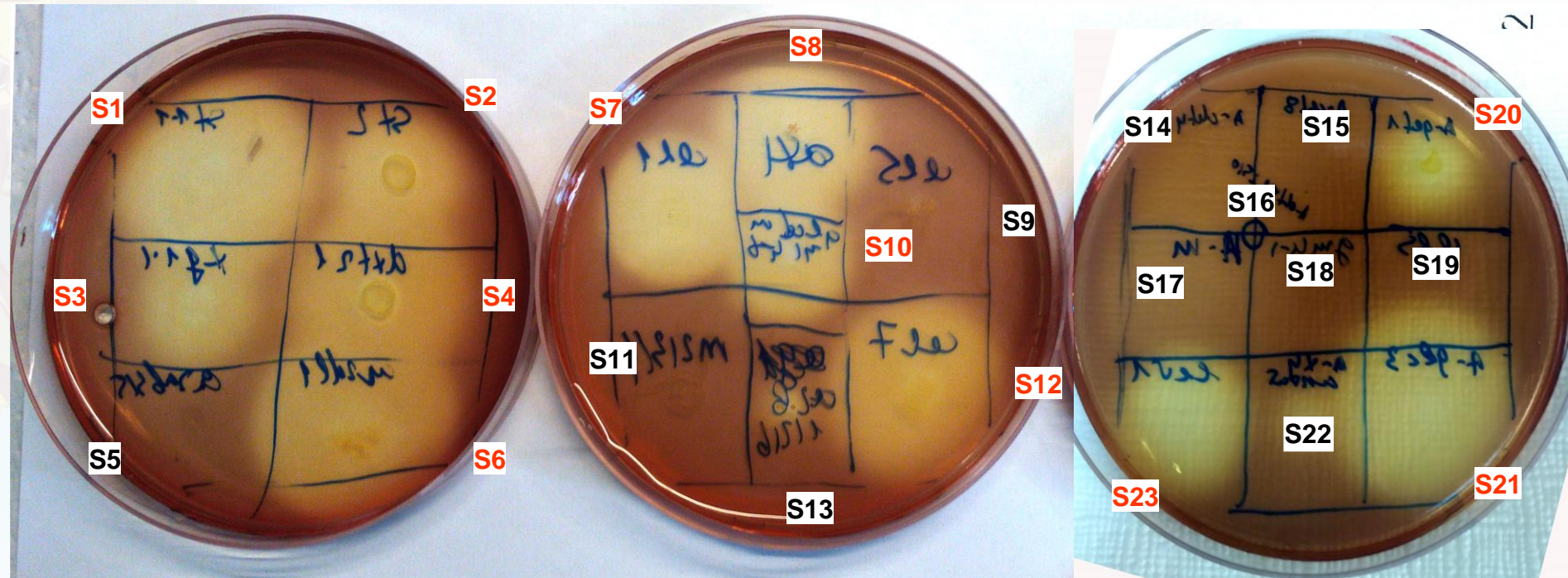
AArc-LBj



AArc-L2

Lipase activity in colonies of natronoarchaea





Soda lake natronoarchaea

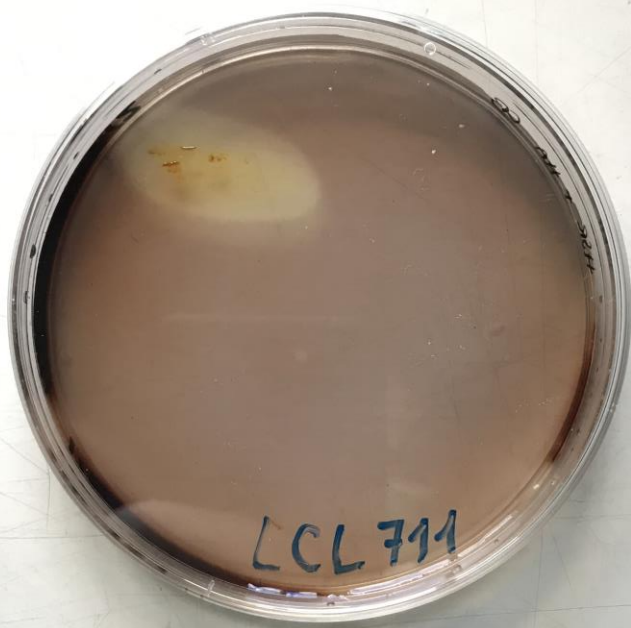
- S1:** AArc-St1-1 (amylolytic)
- S2:** AArc-St2 (amylolytic)
- S3:** AArc-xg1-1 (xyloglycan)
- S4:** AArc-dxtr1 (dextran)
- S5:** AArc-arb3/5 (arabinan)
- S6:** AArc-curd11 (curdlan)
- S7:** AArcel1 (*Natronolimnobius*; cellulo-xylan)
- S8:** AArc-ax1 (*Natronolimnobius*; arabinoxylan)
- S9:** AArcel5 (*Natronobiforma*; cellulo-xylan)
- S10:** AArc-glctm3/4/8 (*Natronococcus*; glct-mannan)
- S11:** AArc-m2/3/4 (mannan-cellulo)
- S12:** AArcel7 (chitin)
- S13:** AArc-arb1/2/6 (*Natronolimnobius*; arabinan)

S13

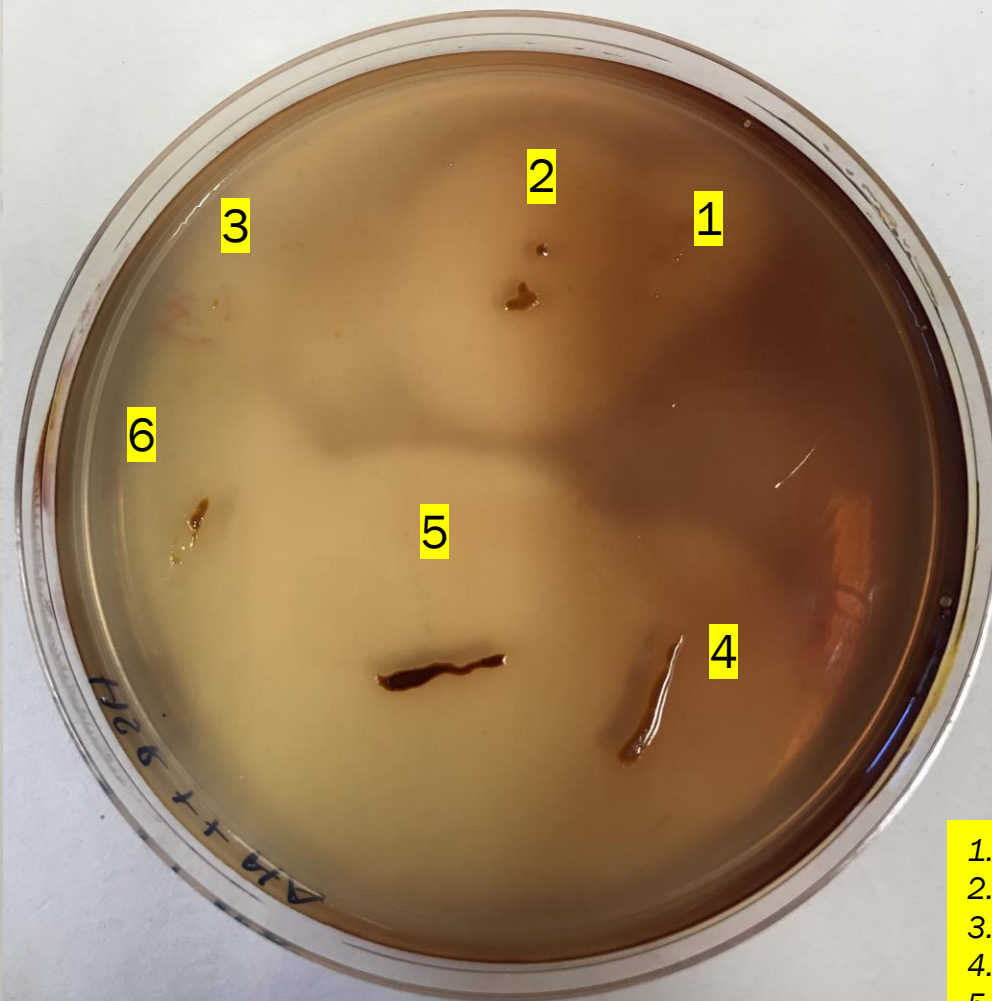
- S14:** AArc4 (*Natrarchaeobius chitinivorans*)
- S15:** AArc8 (*Natrarchaeobius chitinivorans*)
- S16:** AArc-SI (*Natrarchaeobius chitinivorans*)
- S17-19:** Mannan-utilizing natronoarchaea
- S20:** AArc-glct1 (galactan)
- S21:** AArc-glc3 (*Natronorubrum tibitense*; glycogen)
- S22:** AArc-X4 (*Halomicrobium* sp; cellulo)
- S23:** AArc-lev1 (levan)[~AArc-St1-1]

Only **AArcel7** showed moderate growth with Hyl in liquid culture without ye. But it faded after second transfer

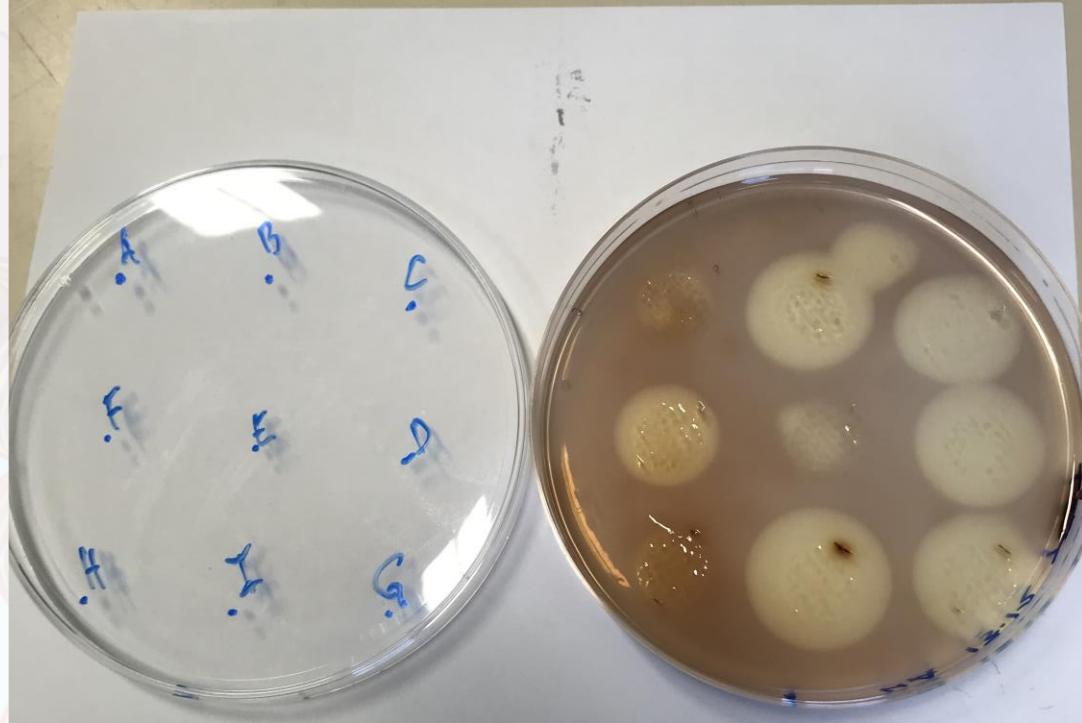
Colony growth: **S1, S2, S3, S4, S6**(weak), **S7, S8, S9(w), S10, S11, S12, S13; Cl6**



Halorhabdus sp. LCL711



1. *Halomicrobium* sp. LC1Hm
2. *Halorhabdus uthaensis* Siv8X
3. *Halomicrobium* sp. Harcel-Eu2
4. *Halococcoides cellulosivorans* BNX81
5. *Halorhabdus* sp. RMX81
6. *Halorhabdus uthaensis* RMX62



- A. *Haloferax lucertense* SVX82
- B. *Halosimplex* sp. HArceI2
- C. *Halomicrobium* sp. HArceI3
- D. *Halobacteria* (new genus) H-hyl
- E. *Haloferax alexandrinus* BNX82
- F. *Halorhabdus* sp. KCL-HA6
- G. *Halococciodes* sp. BariCL
- H. *Haloferax* sp. SVXCL
- I. *Halorhabdus* sp. KCL5

Hyaluronidase activity by agar-diffusion test of haloarchaeal cultures grown with cellobiose + hyaluronate for one week. Fraction of 50 μ l of supernatant was placed on filter discs and incubated on the plates with hyaluronic acid (400 mg l⁻¹) at pH 7.5, 4 M total Na⁺, 37 °C, 24h;

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- Three lipases have been identified by **CSIC**, when screening two fosmid libraries made at BANGOR using tributyrin, olive oil, egg yolk, cocoa and coconut
 - 2 clones from D2 library (bone [turkey femur]-degrading microbiome; 11.12.2017; Byfjorden (60,238185N, 5,181210E) were found positive for tributyrin, egg yolk and cocoa
 - D2 pCCFOS fosmid library has a titre of 9000 clones max.
 - 1 clone from I3 library (bone [cow tibia]-degrading microbiome; 11.12.2017; Byfjorden (60,238185N, 5,181210E) were found positive for tributyrin, egg yolk, olive oil and cocoa
 - I3 pCCFOS fosmid library has a titre of 2000 clones max.

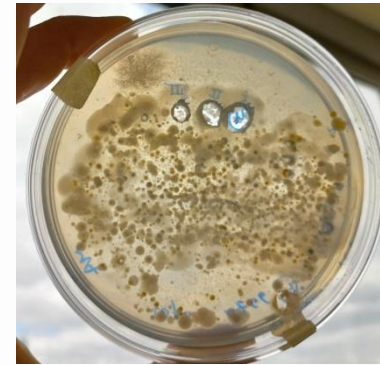
Previously
reported

Task 3.2 Sampling extreme environments for generating new microbial bio-resources

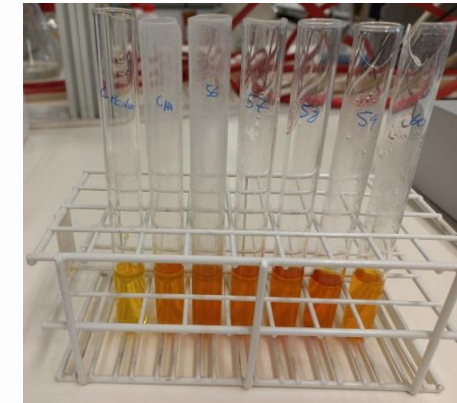
- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

New samples since June (M12-M18)

Graciosa island,
the Azores, PT



Samouco
salterns, PT

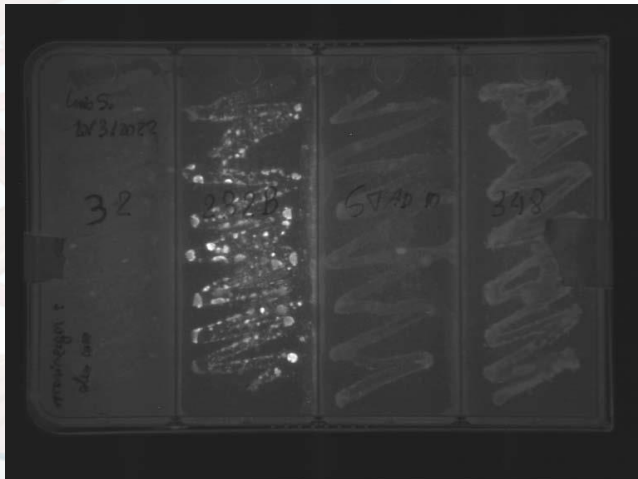


Task 3.1 Exploitation of the FuturEnzyme bio-resources collections

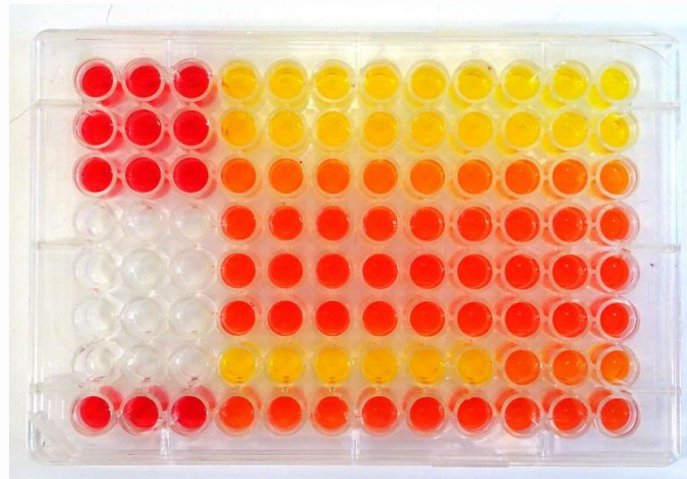
- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

Lipases/Esterases

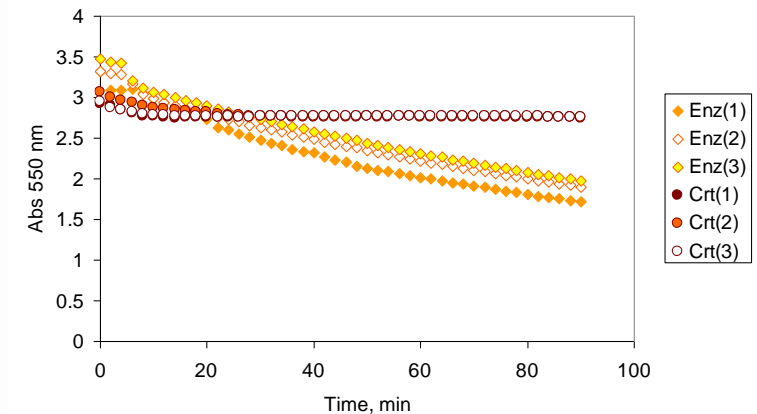
Bacterial collection	Tributyrin or Tween 80	Cononut oil	Palm oil	Olive oil
Existing	40*	5	0	6
FuturEnzyme (new)	7	12	1	16



Screening isolates with lipase activity



Effect of olive oil concentration during bacterial growth on lipase activity

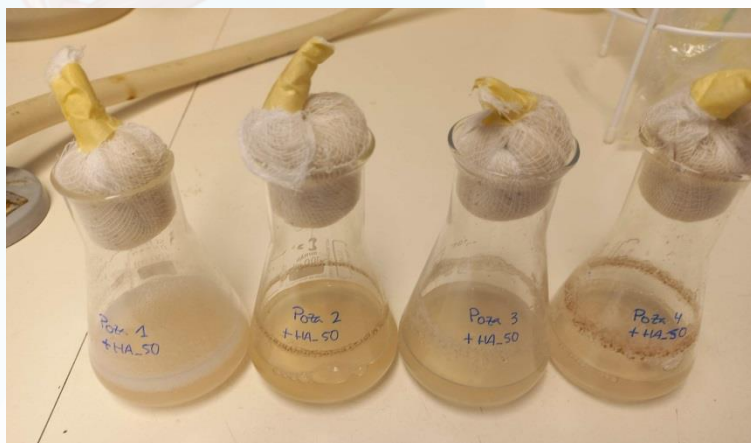


Task 3.1 Exploitation of the FuturEnzyme bio-resources collections

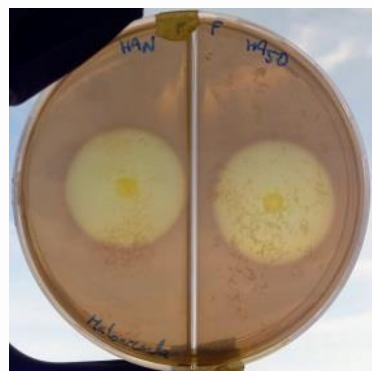
- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

Hyaluronidases

Bacterial collection	HA (Hyacare)	HA5 (Hyacare)
Existing	2*	2*
FuturEnzyme (new)	14	14

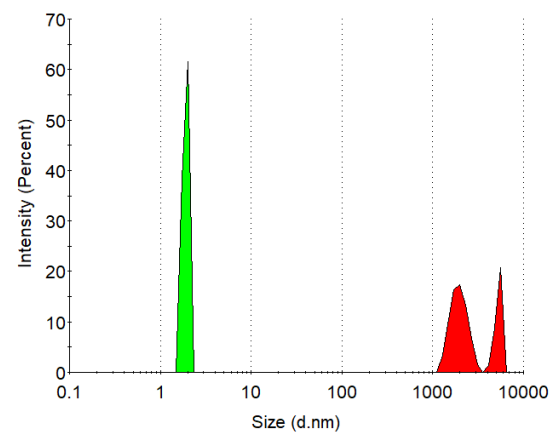


Enrichment cultures with HA and HA50



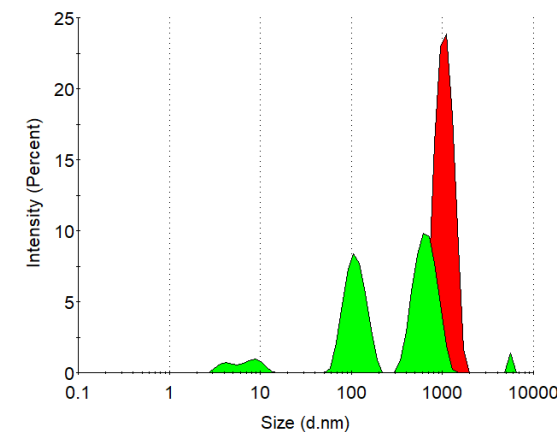
Screening

Laser light scattering



IST_759_HA_0h (red) IST_759_HA_7 days (green)

Stutzerimonas stutzeri



IST_758_HA50_0h (red) IST_758_HA50_7 days (green)

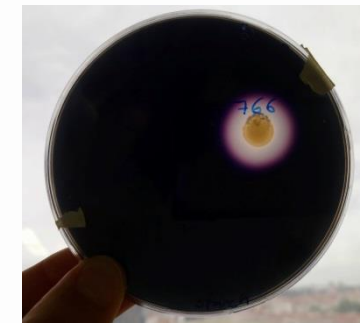
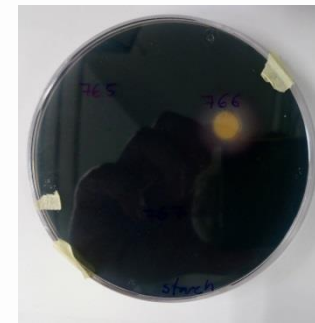
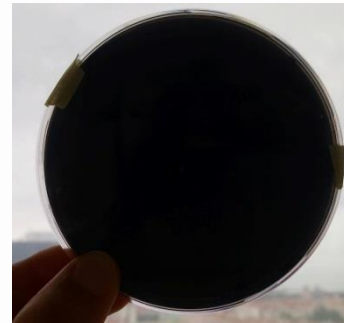
Microbacterium oxydans

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections

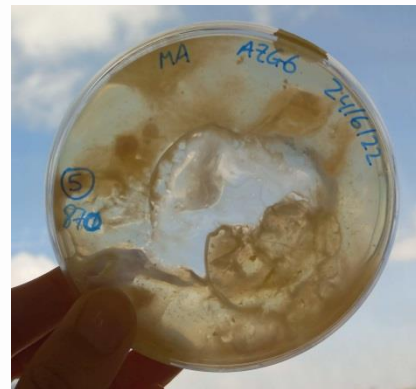
- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

Amylases

Bacterial collection	Starch
Existing	48*
FuturEnzyme (new)	7



Agarases



Task 3.1 Exploitation of the FuturEnzyme bio-resources collections

- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

Other activities

Bacterial collection	Protease	Inulinase	Transaminase
Existing	48*	46*	7*

* Mostly *Bacillus* sp.; presented in MS9

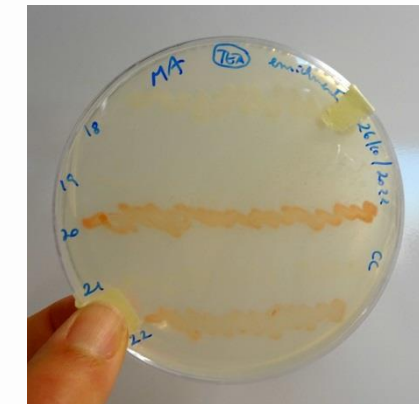
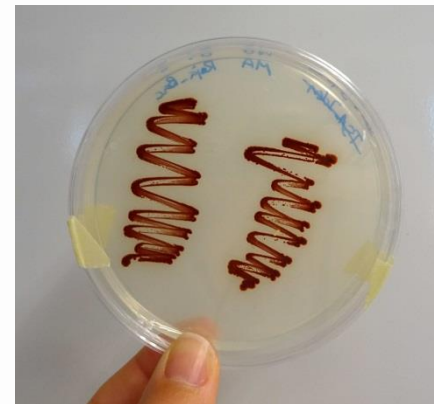
Production of interesting compounds



Melanin by
Neophaeothea triangularis



Prodiginines by
Serratia sp.



Carotenoids by several species

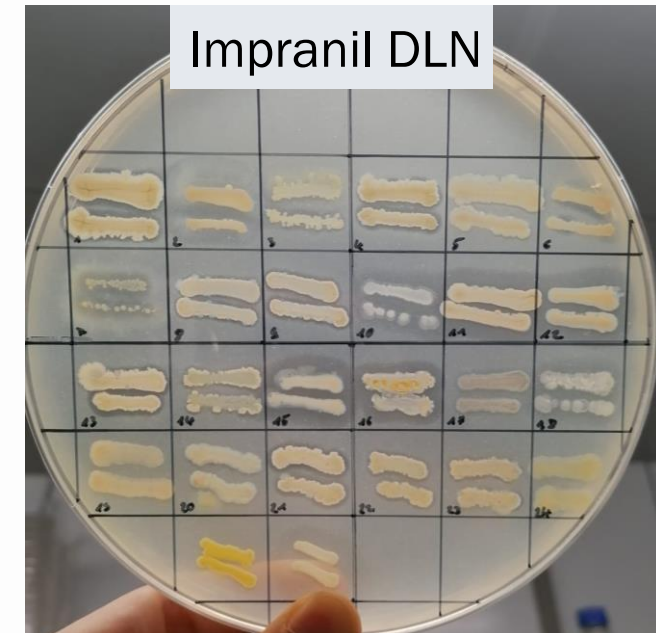
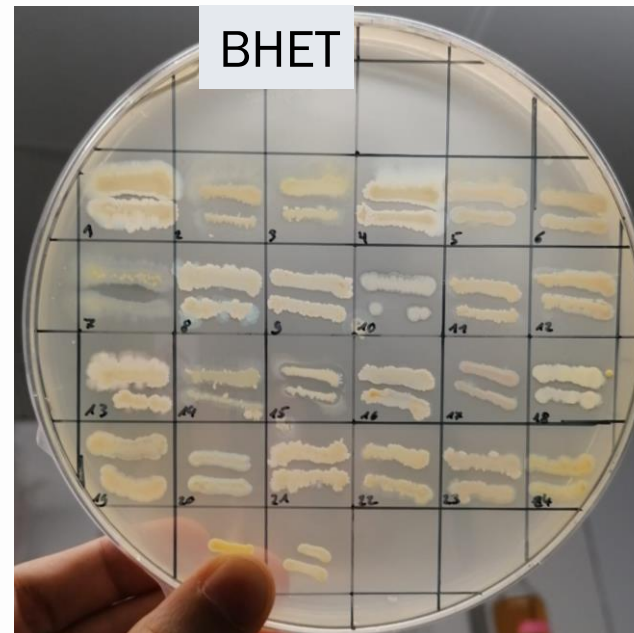
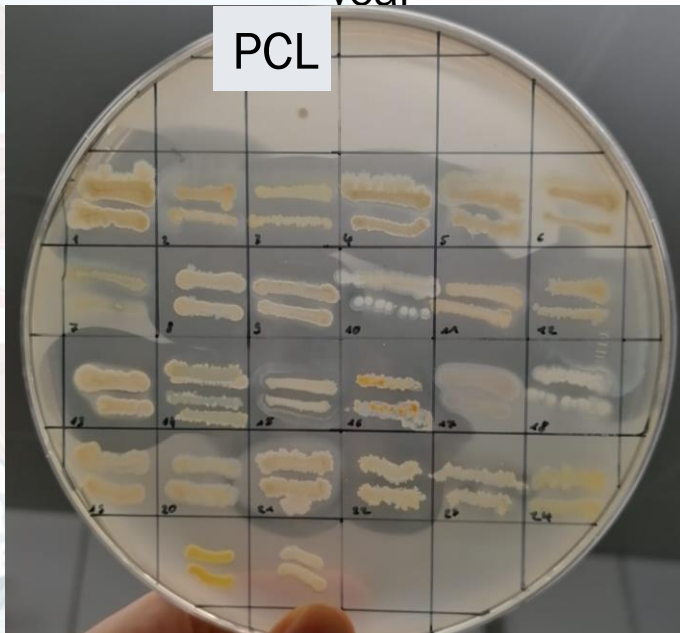
Enrichments for polymer-degrading microorganisms



2005
2012



Enriched in M9 medium, anaerobic + aerobic, 28°C, over up to 1 year



Event name/references

Enrichments for polymer-degrading microorganisms



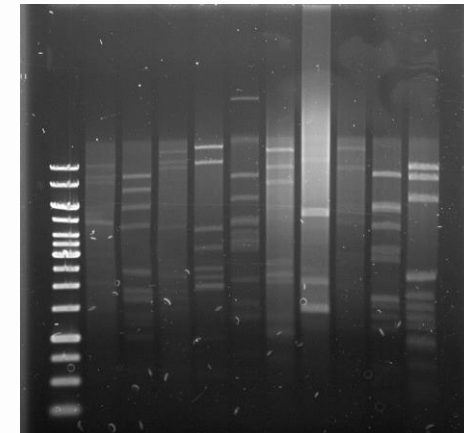
16 S Sequencing results of isolated single strains

1	<i>Bacillus aearus</i>
2	<i>Stutzerimonas stutzeri</i> strain ATCC 14405
3	<i>Bacillus</i> sp.
4	<i>Pseudomonas aeruginosa</i> PA14
5	<i>Achromobacter</i> sp.
6	<i>Pseudomonas mendocina</i> strain NEB698
7	<i>Bacillus altitudinis</i>
8	<i>Pseudomonas plecoglossicida</i>
9	<i>Pseudomonas putida</i>
10	<i>Pseudomonas mendocina</i>
11	<i>Pseudomonas putida</i> strain KT2440
12	<i>Priestia megaterium</i>
13	<i>Pseudomonas proteolytica</i>
14	<i>Bacillus altitudinis</i>
15	<i>Stutzerimonas stutzeri</i> strain ATCC 14405
16	<i>Pseudomonas indoloxydans</i>
	<i>Acinetobacter</i> sp. NyZ410

Fosmid library of *Rhodococcus fascians*



photo: DMSZ



30-37 kb inserts

600 clones

Substrate	Degrading Colonies
DLN	2
BHET	2
PCL	1
TBT	many

to be verified...

Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

- 34 lipases have been identified by CSIC, when screening 5 fosmid libraries made at BANGOR using tributyrin, olive oil, egg yolk, cocoa and coconut

# Site-library	Nr. Clones	Nr of positives
D2 library (bone [turkey femur]-degrading microbiome; 11.12.2017; Byfjorden (60,238185N, 5,181210E)	9000	2
I3 library (bone [cow tibia]-degrading microbiome; 11.12.2017; Byfjorden (60,238185N, 5,181210E)	2000	1
MedSea clone library (Ancona port, Italy, 43 ° 37'N; 13 ° 30'15"E)	10300	7
Acid mine drainage system (Spain; 43 ° 15'47"N, 5 ° 46'9"W)	11600	13
TB (Thermophilic Bacteria) (mix genomes)	11800	11
TOTAL	44700	34



Previously
reported

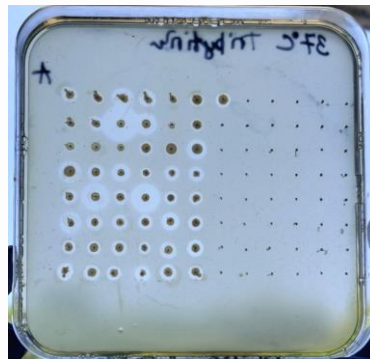
Task 3.1 Exploitation of the FuturEnzyme bio-resources collections (M1-M24)

Screening of Ischia hot vents BBP-enrichment fosmid library (Ischia, Italy) (10000 clones max) identified:

- 12 fosmid clones positive for tributyrin, 2 fosmids clones positive for tributyrin, coconut oil, palm oil
- 16 fosmid clones with amylase activity for starch
- 3 fosmid clones with laccase activity for syringol and 3 fosmid clones with protease activity for skim milk

Screening of soil fosmid library (9000 clones max) identified:

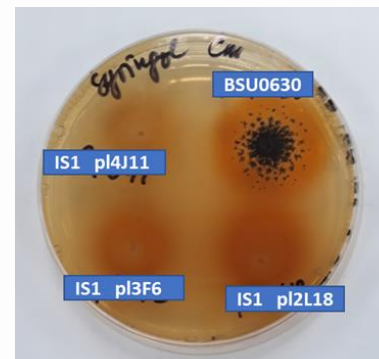
- 16 fosmid clones were positive for tributyrin



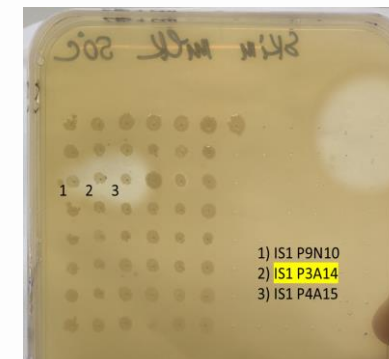
tributyrin



starch



syringol



skim milk

Previously reported

Task 3.2 Sampling extreme environments for generating new microbial bio-resources M6-M30

- Sampling activities are planned to generate new isolates and sequences to feed WP2/WP4 ***in progress***

Milestone 11: ***"The first sampling campaign completed"***

Task 3.2 Sampling extreme environments for generating new microbial bio-resources M6-M30

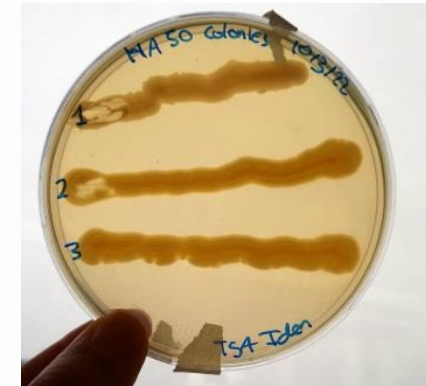
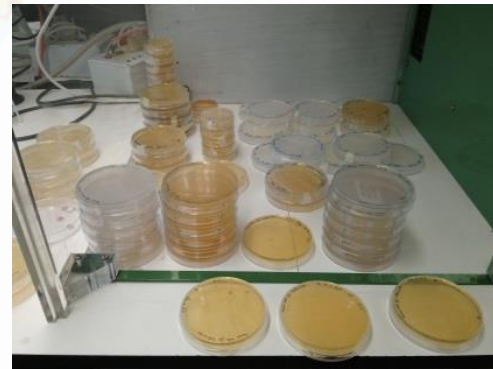
- Sampling activities are planned to generate new isolates and sequences to feed WP2/WP4 **in progress**



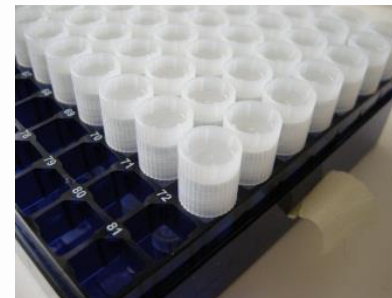
Task 3.2 Sampling extreme environments for generating new microbial bio-resources

- sampling for new bio-resources; identifying novel microbes and enzyme activities; screening our microbial collection for efficient enzymes

Rock pool at Guincho, Portugal



Sherlock® Microbial ID System



FuturEnzyme



Screening

Previously
reported



Task 3.3 Next Generation Sequencing for target enzymes

M1-M36

- *Thermoleophilum album* – thermophilic, obligate hydrocarbonoclastic, high-GC Gram+. Genome sequenced at Bangor
26 genes were cloned, **5** soluble esterases purified, **2** lipases were found active with C14-C18 pNp-esters

Previously
reported

Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36



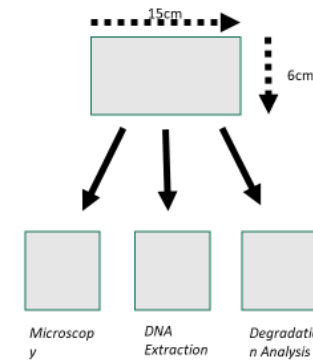
Sequences available for enzyme mining:

- 39 treatments/timepoints of microbiomes of colonisers of plastics (HDPE, LDPE, PP, PE and PET) in a transition:
 - wastewater treatment plant effluent
 - river water
 - brackish water
 - seawater (mesocosm) at the School of Ocean Sciences, Bangor University

Sequenced by Bangor (NERC 'Plastic Vector' project)
>250 Gb data available for enzyme mining in FuturEnzyme

Previously
reported

Water	Time
Effluent	24 Hours
Fresh	16 Hours
Brackish	18 Hours



Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36

Menai Straits surface seawater an the St George Pier, School of Ocean Sciences, Bangor University

Lignin enrichment (shotgun metagenome sequenced at Bangor)

18 genes from 6 enzyme families with highest coverage were cloned

Previously
reported



Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36

- 30 treatments/timepoints of microbiomes from anaerobic bioreactors set up from Landfill in Penhesgyn Recycling Centre, Anglesey Natural Resources Wales (co-participant) 200 Gbp sequencing data
 - 11** amylases (most-abundant in metagenomic reads) (s. overleaf) cloned, **5** expressed soluble, **1** found active with starch substrates
 - 8** Hyaluronidase cloned, **6** expressed soluble.

Previously
reported



Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36

Bangor: Sequencing isolates for UDUS. Sequencing strains of rhodococci purchased at DSMZ

Your Samples

Barcode ?	Type	Coverage ?	Customer's Ref	Taxon	Current Queue (Status)
238336	WGS_DNA	30x	Sample1	Spirosoma endbachense	Bioinformatics
238337	WGS_DNA	30x	Sample2	Raoultella ornithinolytica	Bioinformatics
238338	WGS_DNA	30x	Sample3	Raoultella ornithinolytica	Bioinformatics
238339	WGS_DNA	30x	Sample4	Pseudomonas knackmussii	Bioinformatics
238340	WGS_DNA	30x	Sample5	Microbacterium profundii	Bioinformatics
238341	WGS_DNA	30x	Sample6	Escherichia fergusonii	Bioinformatics
238342	WGS_DNA	30x	Sample7	Phenylobacterium falsum	Bioinformatics
238343	WGS_DNA	30x	Sample8	Erythrobacter pelagi	Bioinformatics
238344	WGS_DNA	30x	Sample9	Rhodococcus fascians	Bioinformatics
238345	WGS_DNA	30x	Sample11	Rhodococcus rhodochrous	Bioinformatics
238346	WGS_DNA	30x	Sample12	Rhodococcus rhodochrous	Bioinformatics
238347	WGS_DNA	30x	Sample13	Rhodococcus sp.	Bioinformatics
238348	WGS_DNA	30x	Sample14	Rhodococcus sp.	Bioinformatics
238349	WGS_DNA	30x	Sample15	Rhodococcus rhodochrous	Bioinformatics

UDUS

Bangor (DSMZ strains)

Complete on November 7, 2022

Task 3.3 Next Generation Sequencing for generating sequences of target enzymes M1-M36

Bangor:
Sequencing of isolates for CNR Messina. Hyaluronidase and other GH-positive.

Your Samples

Barcode ?	Type	Coverage ?	Customer's Ref	Taxon	Current Queue (Status)
233161	WGS_DNA	30x	PG22_01	Halorhabdus	Complete
233162	WGS_DNA	30x	PG22_02	Halorhabdus	Complete
233163	WGS_DNA	30x	PG22_03	Halorhabdus	Complete
233164	WGS_DNA	30x	PG22_04	Halorhabdus	Complete
233165	WGS_DNA	30x	PG22_05	Halorhabdus	Complete
233166	WGS_DNA	30x	PG22_06	Halorhabdus	Complete
233167	WGS_DNA	30x	PG22_07	Halorhabdus	Complete
233168	WGS_DNA	30x	PG22_08	Halorhabdus	Complete
233169	WGS_DNA	30x	PG22_09	Halorhabdus	Complete
233170	WGS_DNA	30x	PG22_10	Vibrio alginolyticus	Complete
233171	WGS_DNA	30x	PG22_11	Vibrio alginolyticus	Complete
233172	WGS_DNA	30x	PG22_12	uncultured gamma proteobacterium	Complete

Haloarchaea
(CNR)

Marine gamma-proteo
(CNR)

Complete on July 1, 2022

Task 3.3 Bio-resources (Bangor) (new)

62 genes synthesized since last meeting in Madrid (in addition to **37** previously synthesized at Bangor) for:

1. 8 glycosyl hydrolases (new HA-candidates from sequenced genomes of **Partner CNR**))
2. **19** Lipases, esterases, laccases, peroxidases-catalases from *Oleiphilus messinensis* ME102^T
3. **9** hydrolases from Parys Mt hyperacidic (pH 1.5) fosmid library
4. **4** MCO, peroxidases from PP and LDPE colonisers
5. **22** LipEst, MCO, peroxidases/catalases from 1-week PET colonisers shotgun metagenome-sequencing data (Menai Strait seawater mesocosm)

Deliverables

List of deliverables

Deliverable Number ¹⁴	Deliverable Title	Lead beneficiary	Type ¹⁵	Dissemination level ¹⁶	Due Date (in months) ¹⁷
D3.1	Bio-resources prepared and exchanged	3 - BANGOR	Other	Confidential, only for members of the consortium (including the Commission Services)	2
D3.2	Standard assays, analytics and calculations for monitoring enzymatic performance	4 - UHAM	Report	Confidential, only for members of the consortium (including the Commission Services)	2
D3.3	Set of 100 best clones, 10 isolates, and 10 enzymes shortlisted for sequencing or transfer to WP2	6 - IST ID	Other	Confidential, only for members of the consortium (including the Commission Services)	10
D3.4	Sequence, activity, and stability datasets from best positive bio-resources	3 - BANGOR	data sets, microdata, etc	Confidential, only for members of the consortium (including the Commission Services)	18
D3.5	Set of new bio-resources to screen or sequence	7 - CNR	Other	Confidential, only for members of the consortium (including the Commission Services)	24
D3.6	Complete set of positive naïve screened enzymes and sequences and their datasets	3 - BANGOR	Report	Confidential, only for members of the consortium (including the Commission Services)	32



Milestones

Milestone number ¹⁸	Milestone title	Lead beneficiary	Due Date (in months)	Means of verification
MS9	First round of functional screens completed	6 - IST ID	6	Materials available – this milestone will attest the realisation of the first screens of available bio-resources.
MS10	First round of sequencing completed	3 - BANGOR	6	Data available – this milestone will attest to the realisation of the sequencing of the first selected bio-resources found to be positive in the screen tests.
MS11	The first sampling campaign completed	7 - CNR	12	Sites data, samples available - this milestone will attest completion of campaigns for sampling new bio-resources with information about sample sites available.

